REMARKS

Claims

Claims 1 and 5-9 are currently amended and claims 2-4 and 10-11 are cancelled.

Claim 1 is amended to incorporate the limitations of dependent claims 2-4, and to modify the antecedent relating to the alkyl ether group in claim 1 so it properly encompasses the later limitations of claim 6. Claim 1 now encompasses an alkyl ether that is any of a saturated, unsaturated or cycloalkyl group and claim 6 is amended such that the alkyl portion of the alkyl ether group is characterized by the formula of C_nH_{2n+1} wherein n is at least 3 and less than 20. Support for these amendments in claims 1 and 6 is found in the application on p. 2, lines 15-16 and p. 5, lines 25-26. Applicants therefore respectfully submit that there is now a proper antecedent basis in claim 1 for the elements in claim 6.

Claims 5-9 are herein amended to make them consistent with amended claim 1, wherein claim is now directed to a pharmaceutical formulation rather than a compound. Therefore, claims 5-9, which depend from claim 1, are all now directed to a pharmaceutical formulation comprising a cytoprotection effective dose.... Support for these amendments is found in the application on p. 2, lines 26-29 and p. 3, lines 25-28 and original claim 11. Applicants submit that with these amendments, no new matter has been added.

Request for complete reference to J. Bone Miner Res. filed in IDS paper no. 10

The examiner has requested the complete reference for the above-reference abstract submitted with the IDS of November 4, 2002. Unfortunately, the abstract is the only reference available. This is the published abstract to accompany the oral presentations presented at the Annual Meeting of the American Society for Bone and

Mineral Research in September of 2001. A list of Dr. Pierce's publications from the website at the University of Louisville in Kentucky (see Appendix A, attached hereto) shows that no accompanying follow-up journal article was published on this topic.

Therefore, Applicants are unable to provide a complete reference to this abstract as it is believed none exists.

Specification, 35 U.S.C. § 112, para. 1

The Examiner has entered a 35 U.S.C. § 112, para. 1 citation stating that the specification does not meet the standards of being written in "full, clear, concise and exact terms" because it is "replete with terms which are not clear, concise and exact." See Office Action, p. 2. The Examiner requests that the specification be checked to correct any errors. Applicants have done this, and in the interest of efficiency, are submitting herewith a Substitute Specification with wherein spelling and other minor typographical errors have been corrected.

The Claims as Amended are not Indefinite Under 35 USC § 112, para. 2

Claims 1 and 5-9 are herein amended from "A cytoprotective compound" to "A pharmaceutical compound, comprising *a cytoprotection effective dose....*" Applicants respectfully submit that the claims are not indefinite, as amended. "A cytoprotective effect" is defined in the application on p. 4, lines 31-32. Further, the application discusses that "experimental models for measuring cytoprotection" are known in various cell lines (p. 6, lines 23), and provides references for such (id.) and a description in Example 2 (p. 16). A discussion of the use of effective doses of compounds described in the application to treat chronic degenerative disorders is found on p. 6, lines 29-30 and

what might constitute and effective dose in any given formulation is found on p. 9, lines 22-30, particularly lines 29-30.

The Examiner also pointed out the presence of an improper antecedent basis in claim 6 relative to claim 1. As herein amended, Applicants respectfully submit that claim 6 has proper antecedent basis for all the elements, and as such is definite.

Finally, the Examiner has also stated that it is "unclear what is the meaning of 'comprising' in [the] claims." See Office Action, p. 3, lines 4-5. Claim 1 refers to a pharmaceutical formulation "comprising" a compound having an estrogen ring structure. It is well understood that use of the phrase "comprising" as it is used in claim 1 means that, in addition to the elements recited in claim 1, the formulation may also include other components. For example, the formulation may include an excipient, solvent, or other agent that aids in delivery, or may include an additional active ingredient. As such, Applicants respectfully submit that the claims are not indefinite and request withdrawal of the § 112, para. 2 rejection.

35 U.S.C. § 112, para. 1, Enablement

The Examiner has rejected claims 1-11 for lack of enablement (see Office Action, p. 3) asserting that the specification, while being enabling for estradiol, does not reasonably provide enablement for any other steroid. *Id.* As herein amended, claims 1 and 5-10 are directed to a pharmaceutical formulation comprising "a four ring estrogen structure having a first end and a second end wherein a phenol group is located at the first end and a D carbon ring having an alkyl ether functional group on carbon 17 is located at the second end. Applicants respectfully submit that as amended, claims 1 and 5-10 more particularly point out that the claimed formulation comprises a compound having an

estrogen ring structure and so the claim invention is thus enabled by the specification, does not encompasses unpredictable art as estrogens are a well-studied and well-defined art, are not overly broad, has sufficient guidance in the specification for how to practice and use the claimed invention, provides sufficient working examples, and does not require undue experimentation. Applicants therefore respectfully request withdrawal of the enablement rejection.

Judicial Doctrine of Double Patenting

Claims 1-11 are rejected under the judicially created doctrine of double patenting over claims 1-29 of '601 patent. The Examiner believes that, if allowed, the claims of the present application would improperly extend the patentee's right to exclude, which was already granted in the '601 patent. The office action appears unclear as to the nature of the rejection based on the '601 patent. On the one hand, the basis for rejection is said to be the judicially created doctrine of double-patenting. See Office Action, p. 2, next to last paragraph. This doctrine applies only when the subject matter *claimed* herein would have been obvious in light of the subject matter *claimed* in the '601 patent. As stated in MPEP 804, judicially created double-patenting is "grounded in public policy" and is "primarily intended to prevent prolongation of the patent term by prohibiting *claims* in a second patent not patentably distinct from *claims* in a first patent."

On the other hand, in the next sentence the same reference is said to have "fully disclosed" the subject matter of the present application, apparently a 35 U.S.C. § 102(b) rejection (a circumstance that is distinct from the judicially created doctrine of double-patenting). The last sentence on p. 2 takes a third position, namely that the same invention as claimed herein was patented in the '601 patent, apparently a statutory

double-patenting rejection under 35 U.S.C. § 101. Although it is logically possible for the second and third rejections to co-exist, all three of these rejections cannot logically exist together.

A further rejection also is made on the basis of the '601 patent, namely that is renders obvious the subject matter of the claimed invention here under 35 U.S.C. § 103(a). See Office Action, p. 6.

Regardless of the particular rejection, Applicants respectfully submit that the subject matter of the claimed invention is not anticipated by the '601 patent (thus there can be no 102(b) rejection), is not the same as the subject matter of the '601 patent (thus there is no basis for a statutory double-patenting rejection), contains claims that are patentably distinct from claims previously patented in a prior application (thus there is no basis for an obviousness-type double patenting rejection under the judicially created doctrine of double-patenting) and is not obvious in light of Simpkins et al. (thus there is no basis for a 103(a) rejection).

No Basis for a Statutory Double-Patenting Rejection Under 35 U.S.C. § 101 or an Anticipation Rejection Under 35 U.S.C. § 102(b).

The present claims are directed to a pharmaceutical formulation that comprises a cytoprotection effective amount of a compound having an estrogen ring structure and an alkyl ether on carbon 17 of the D ring, wherein the alkyl portion of the alkyl ether is any of a long chain saturated alkyl, a long chain unsaturated alkyl, or a cycloalkyl group. The '601 patent discloses estrogen compounds that have cytoprotective activity, but neither discloses nor claims the specifically modified compounds of the present claims. The '601 patent does disclose a generic formula similar to that claimed in the present

invention, but no disclosure of a long-chain alkyl ether, saturated or unsaturated, or a cycloalkyl ether, for the R₂ group at carbon 17 of the D ring is provided. As stated in MPEP § 804 (II) (A), a reliable test for double-patenting ... is whether a claim in the application could be literally infringed without literally infringing a corresponding claim in the patent. In re Vogel, 422 F.2d 438, 164 ... (CCPA 1970). Is there an embodiment of the intention that falls within the scope of one claim, but not the other? ... For example, the invention defined by a claim reciting a compound having a "halogen" substituent is not identical to a claim reciting the same compound except having a "chlorine" substituent in place of the halogen because "halogen" is broader than "chlorine." In the present case, this example applies because the estrogen compounds provided in the '601 patent are for conferring *neuro* protection on a population of cells, whereas the presently claimed subject matter is for a pharmaceutical formulation comprising a "cytoprotection effective dose" of an estrogen compound. First, cytoprotection is broader than neuroprotection, so the claimed subject matter is not the same. Second, it would be possible to infringe the claimed invention in the instant application without infringing the claimed invention in the '601 patent. The '601 patent claims a method for conferring neuroprotection with two steps – (i) providing an estrogen compound and (ii) administering the compound whereas the instant application claims a pharmaceutical formulation comprising a cytoprotection effective dose of an estrogen compound. A method claim and a product claim are not synonymous. Even if one were to actually use the pharmaceutical formulation of the instant application, as disclosed on pp. 2-3 in the fourth embodiment of the specification of the instant application, the population of cells or tissues that can be targeted to retard development of a degenerative condition includes

"... stem cells, blood cells, epithelial cells, ... muscle tissue cells, ... bone cells, skin cells, reproduction tract cells, and urinary tract cells." Treatment of such cells with compounds of the instant application would not infringe a claim for "A Method for conferring neuroprotection in a population of cells in a subject..." because none of those cells are neural cells. Thus, there is no basis for a statutory double-patenting rejection under 35 U.S.C. § 101 or an anticipation rejection under 35 U.S.C. § 102(b). No Basis for a Non-Statutory Obviousness-Type Double Patenting Rejection or for an Obviousness Rejection Under 35 U.S.C. § 103(a)

As noted above, the judicially created double-patenting is "grounded in public policy" and is "primarily intended to prevent prolongation of the patent term by prohibiting *claims* in a second patent [application] not patentably distinct from *claims* in a first patent." As amended herein, the claims in the instant application are patentably distinct from the claims in the '601 application.

The analysis employed in an obviousness-type double patenting determination parallels the guidelines for a 35 U.S.C. §103(a) rejection (see MPEP § 804(II)(B)(1)). Therefore, both the obviousness-type double-patenting rejection and the §103(a) rejection will be discussed together. If the presently claimed invention is not obvious under 35 U.S.C. §103(a) then an obviousness-type double-patenting rejection is not appropriate.

There are three requirements for a *prima facie* case of obviousness: 1) some suggestion or motivation, in the reference itself or the knowledge generally available in the art, to modify the reference to arrive at the claimed invention; 2) there must be a reasonable expectation of success; and 3) the reference must teach or suggest all the claim limitations. See MPEP § 706.02(j). In the instant case, there is no *prima facie* case

as there is no suggestion in Simpkins et al., or the knowledge generally available in the art, to particularly use an alkyl ether, whether long chain saturated, unsaturated, or cycloalkyl at position 17 of the D ring of the estrogen compound (at the R₂ position in Simpkins et al.).

The '601 patent provides a variety of possible substituents at the R_2 position, from -OH to benzoate, from ethynyl $-\alpha$ to ketal, from triethyl ammonium salt to sodium phosphate to (see Simpkins et al., Figs. 9A and 9B and claims 4-5 and 22, among others). Some of the possible R_2 groups in the '6-1 patent are esters, others are salts, some are hydroxyls, some are aromatic, some are branched, some are linear, some are short chains, some are long. There is no particularity in the possibility for R_2 in the '601 patent, and no suggestion in the '601 patent, or the knowledge generally available in the art, that long chain alkyl ethers or cycloalkyl ethers are particularly desirable for providing cytoprotection in cells.

As one skilled in the art would know, short chain alkyl ethers are either completely water miscible (methoxymethane) or partly water miscible (ethoxyethane) (see Chapter 9, Alcohols and Ethers, p. 291, in Organic Chemistry, 2nd Edition, Volhardt, K., et al. (1994), W. H. Freeman and Co., New York – attached herewith as Appendix B). Therefore, as one increases the length of the alkyl chain, or the overall number of carbons, the hydrophobicity increases. Further, since each carbon bond in an alkyl chain has free rotation about the carbon, as chain length increases, the degrees of freedom for conformational flexibility also increases. The alkyl groups required in the presently claimed position at carbon 17 in the D ring are all hydrophobic and at least 3 carbons in length. There is no such distinction for the possible R₂ groups in the '601 patent, and no

suggestion that the R_2 groups be limited to hydrophobic groups, 3-20 carbons, or to anything at all in particular.

In contrast, it was surprisingly found that certain exemplary embodiments of estrogens modified at carbon 17 of the D ring can have 10-fold greater cytoprotection in a variety of cells, not just neuronal cells, relative to the core estrogenic compound from which the particular modified estrogen was derived, as long as the modified estrogen obeys the claimed limitations – i.e. there is an alkyl ether at carbon 17 of the D ring which is either a long chain saturated alkyl, long chain unsaturated alkyl, or cycloalkyl ether group. In addition, it was surprisingly found that substituting with either too large a functional group or too small a functional group at the 17 position (R₂ in the 601 patent) can dramatically affect the cytoprotection capability of such modified estrogen compounds. See application, p. 16, line 14-p. 17, line 12. None of these insights are disclosed in the cited reference, nor is any specific compound that is claimed in the instant application, and the cited reference does not suggest such making such modifications to the estrogen compounds disclosed therein. In short, nothing in the cited '601 patent or the knowledge generally available in the art reveals the complexity of the relationship of cytoprotection properties and the nature of the substituent at the 17 position of the D ring. Therefore, the first requirement of a prima facie case – suggestion or motivation to modify – is not present.

In addition, although the generic formula for estrogen disclosed in the '601 patent encompasses the specific compounds claims in the instant application, there is nothing to suggest in the '601 application that one could or should modify the R₂ substituent to arrive at the specific substituent claimed in the instant application. As stated in MPEP

2144.08(II), "The fact that a species or subgenus is encompassed by a prior art genus is not sufficient by itself to establish a prima facie case of obvious ness" quoting from In re Baird, 16 F/3rd 380, 382 (Fed.Cir. 1994) and "The fact that a claimed compound may be encompasses by a disclosed generic formula does not by itself render that compound obvious" quoting from In re Jones, 958 F.2d 347, 350 (Fed.Cir. 1992). the '601 patent does not disclose each element of the claims in the presently claimed invention. Claims 1 and 5-9 of the instant application require an alkyl ether substituent on carbon 17 of the D ring, wherein the alkyl portion is either a long-chain saturated alkyl group, a long-chain unsaturated alkyl group, or a cycloalkyl group. Nothing in the '601 patent discloses that the R_2 position could be, or more importantly, should be, a long-chain alkyl ether, whether saturated or unsaturated, or a cycloalkyl ether. As stated above, the claimed estrogen compounds having the disclosed particular substituents at position 17 of the D ring have been found to convey surprisingly increased cytoprotection, in general, on a wide-variety of cells relative to the generic estrogen compounds disclosed in the 601' patent having non-particular substituents at position 17 of the D ring, which are disclosed to confer cytoprotection only to neuronal cells. See p. 17, lines 3-13 of the present application, which describes that this discovery was unexpected and led to the present definition of n to be at least 3 and less than 20. This element of the present claims was an unexpected and nonobvious discovery over the disclosure of the '601 patent. Thus, the claims are patentable in light of Simpkins '601.

For the above reasons, Applicants respectfully submit that the Examiner has not established a *prima facie* case of obviousness. Therefore the obviousness-type double patenting rejection and the §103(a) obviousness rejection should be withdrawn.

No Basis for a Novelty Rejection Under 35 U.S.C. §102(b)

The Examiner alleges that claims 1-6 (as herein amended, claims 1, 5 and 6) are anticipated by compound RN 119309-39-6 of Levchenko et al., which has the substituent –OBu-i on carbon 17 of the D ring. As herein amended, claims 1, 5 and 6 refer to a pharmaceutical formulation comprising a compound having an estrogen ring structure. As the cited art refers to estrogen impurities, not pharmaceutical formulations comprising estrogen compounds, it does not disclose the claimed invention of a pharmaceutical formulation

CONCLUSION

For the reasons set forth above, it is respectfully submitted that all pending claims are in condition for allowance. Reconsideration of the claims and a notice of allowance are therefore requested.

Applicant submits herewith a Petition for a three- month extension of time, along with the requisite fees. Applicants believe that no additional fees are required. If, however, additional fees are required for the timely consideration of this response, Applicants authorize the Commissioner to charge deposit account number 19-4972 for any additional fees that may be required for the timely consideration of this application.

Date: August 11, 2003

Respectfully submitted,

Barbara J. Carter, Ph.D. Registration No. 52,703

Attorney for Applicants

Bromberg & Sunstein LLP 125 Summer Street Boston, Massachusetts 02110-1618

Tel: (617) 443-9292 Fax: (617) 443-0004



PubMed Bibliography for WM Pierce with Abstracts
PEER REVIEWED RESEARCH PUBLICATIONS
BOOK CHAPTERS, REVIEWS, PATENTS
ABSTRACTS AND PRESENTATIONS

×

PEER REVIEWED RESEARCH PUBLICATIONS:

Thongboonkerd, V., J Luengpailan, J Cao, WM Pierce, Jr., J Cai, JB Klein and RJ

Doyle. Fluoride exposure attenuates expression of Streptococcus pyogenes virulence factors. J. Biol. Chem. 277:16599-16605, 2002

Ping, P, C Song, J Zhang, Y Guo, X Cao, RC Li, W Wu, TM Vondriska, JM Pass, XL Tang WM Pierce Jr., and R Bolli. Formation of prootein kinase C(epsilon)-Lck signaling module confers cardioprotection. J Clin Invest. 109:499-507, 2002.

Molestina, RE, JB Klein, RD Miller, WM Pierce Jr., JA Ramirez and JT Summersgill. Proteomic analysis of differentially expressed Chlamydia Pneumoniae genes during persistent infection of Hep-2 Cells. Infection and Immunity 70:2976-2981, 2002.

Castegna, A M Aksenov, M Aksenova, V Thongboonkerd, JB Klein, WM Pierce, R Booze, WR Markesberry and DA Butterfield. Proteomic identification of Oxidatively Modified Proteins in Alzheimer's disease Brain. Part I: Creatine Kinase BB, Glutamine Synthase, and Ubiquitin Carboxy-terminal Hydrolase L-1. In Press, Free Radic Biol. Med.

Gozal, E., D Gozal, WM Pierce, V Thongboonkerd, JA Scherzer, LR Sachleben Jr., S-Z. Guo, J Cai, and JB. Klein. Proteomic Analysis of CA1 and CA3 Regions of Rat Hippocampus and Differential Susceptibility to Intermittent Hypoxia. In Press, J Neurochem

Shoemaker, LR, JJ Miller, WM Pierce, AJ Schurman, TR Welch and WH Bergstrom. Calcium homeostasis during normocalcemic hypercalcuria: A rat model of hypercalciuric Bartter Syndrome. Submitted to J of Am Soc Nephrol, 2002.

Arthur, John M., Visith Thongboonkerd, Janice A. Scherzer, Jian Cai, William M. Pierce and Jon B. Klein. Differential expression of proteins in renal cortex and medulla: A proteomic approach. Submitted to Kidney International 2002

Thongboonkerd, V, JB Klein, WM Pierce, Jr., AW Jevans and JM Arthur. Sodium Loading Changes Urinary Protein Excretion. Submitted to Am. J. Physiol. 2002.

Steele, PS, MM Jumblatt, NB Smith and WM Pierce Jr. Histatin Expression in Human Lacrimal Gland and Tears. Submitted to Invest Ophthalmol Vis Sci, 2002.

Castegna, A., M. Aksenov, V. Thongboonkerd, JB Klein, WM Pierce Jr., R. Booze, WR Markesbery and DA Butterfield. Proteomic Identification of Oxidatively Modified Proteins in Altzheimer's disease brian. Part II: hihydropyrimidinase-related protein 2, enolase and heat shock cognate 71. Submitted to Free Radical Biol Med 2002

Thongboonkerd, V., E. Gozal, LR Sachleben, Jr., JM Arthur, WM Pierce Jr., J. Cai, J. Chao and JB Klein. Proteomic Analysis Reveals Alterations in the Renal Kallikrein Pathway During Hypoxia-Induced Hypertension. Submitted to J Am. Soc. Nephrology [JASN-200204-0311], April, 2002.

Wan TC, J Cai, WM Pierce Jr. and WL Dean. Plasma Membrane Ca-ATPase Isoform 4 (PMCA4) is Phosphorylated on Tyrosine 1176 in Activated Human Platelets. Submitted to J Biol Chem April 2002

Feng, W, J Cai, WM Pierce, Jr., and Z-H Song. Expression of functional CB2

- cannabinoid receptor in Pichia pastoris for purification and mass spectrometric characterization. Submitted to 2002.
- Ping, P, J Zhang, WM Pierce, Jr. and R. Bolli. Functional Proteomic Analysis of PKC? Signaling Complexes Assocaited with Cardioprotection. <u>Circ Res. Jan 19;88(1):59-62, 2001</u>.
- Madhavi J. Rane, Patricia Y. Coxon, David W. Powell, Rose Webster, Jon B. Klein, Peipei Ping, William Pierce, and Kenneth R. McLeish p38 kinase-dependent MAPKAPK-2 activation functions as PDK2 for AKT in human neutrophils. <u>J. Biol. Chem., Vol. 276, Issue 5, 3517-3523, 2001</u>.
- Nerland, D.E., J. Cai, W.M. Pierce, Jr. and F.W. Benz. Covalent binding of acrylonitrile to specific rat liver glutathione-S-transferases in vivo. <u>Chem. Res. Toxicol.</u>, 14:799-806, 2001.
- Fitzpatrick, J.L., S.L. Ripp, N.B. Smith, W.M. Pierce, Jr. and R.A. Prough. Metabolism of DHEA by Cytochromes P450 in Rodent and Human Liver Microsomal Fractions. <u>Arch Biochem Biophys.</u> 2001 May 15;389(2):278-87.
- Thakkar, R.R., O.-L. Wang, M. Zrouga, W. Stillwell, A. Haq, R. Kissling, W.M. Pierce, Jr., N.B. Smith, F.N. Miller and W.D. Ehringer. Docosahexaenoic acid reverses cyclosporin A-induced changes in membrane structure and function. <u>Biochim Biophys Acta Apr</u> 6;1474(2):183-195, 2000.
- Feltzer, R., R.D. Gray, W.M. Pierce and W.L. Dean. Alkaline Proteinase Inhibitor of Pseudomonas aeruginosa: Interaction of Native and N-terminally Truncated Inhibitor Proteins with Pseudomonas Metalloproteinases. <u>J. Biol. Chem., 275(28), 21002-21009, July 14, 2000.</u>
- Li J, T-Y Yen, ML Allende, RK Joshi, J Cai, WM Pierce, Jr., E Jaskiewicz, DS Darling, BA Macher and WW Young, Jr. Disulfide bonds of GM2 synthase homodimers: Antiparallel orientation of the catalytic domains. J Biol Chem 2000 Dec 29:275(52):41476-86.
- Chen, D., L.C. Waite and W.M. Pierce, Jr. In vitro effects of zinc on markers of bone formation. <u>Biological Trace Element Res.</u> 68:225-234, 1999.
- Chen, D., L.C. Waite and W.M. Pierce, Jr. In vitro bone resorption is dependent on physiological concentrations of zinc. <u>Biological Trace Element Res.</u> 61:9-18, 1998.
- Hou Y., WM Pierce Jr. and NA Delamere The influence of ascorbic acid on active sodium transport in cultured rabbit nonpigmented ciliary epithelium. <u>Investigative Ophthalmology & Visual Science</u>. 39(1):143-50, 1998.
- Wu Q., W.M. Pierce, Jr. Cytoplasmic pH responses to carbonic anhydrase inhibitors in cultured rabbit nonpigmented ciliary epithelium. <u>J Membrane Biol</u> 162:31-38, 1998.
- Wu Q., N.A. Delamere and W.M. Pierce, Jr. Membrane associated carbonic anhydrase in cultured rabbit nonpigmented ciliary epithelium. <u>Invest. Ophth. Vis. Sci., 38: 2093-2102, 1997</u>.

- Song, W., W.M. Pierce, Jr., Y. Saeki, R.A. Prough and R.N. Redinger. Endogenous 7-Oxocholesterol is an Enzymatic Product: Characterization of 7α -Hydroxycholesterol Dehydrogenase Activity of Hamster Liver Microsomes. Arch. <u>Biochem. Biophys.</u>, 328: 272-282, 1996.
- Sullivan, D.M., P.W. Feldhoff, R.B. Lock, N.B. Smith and W.M. Pierce, Jr. Characterization of an Altered DNA Topoisomerase IIα from a Mitoxantrone Resistant Mammalian Cell Line which is Hypersensitive to DNA Crosslinking Agents. Int. J. Oncology 7: 1383-1393, 1995.
- Kaysinger, K.K., W.M. Pierce, Jr. and D.E. Nerland. Quantitative analysis of 2-oxoglutarate in biological samples using high performance liquid chromatography with electrochemical detection. <u>Anal. Biochem. 222: 81-85, 1994</u>.
- Sharir, M., W.M. Pierce, Jr. Di Chen and T.J. Zimmerman. Pharmacokinetics, Acid-Base Balance and Intraocular Pressure Effects of Ethyloxaloylazolamide -- A Novel Topically Active Carbonic Anhydrase Inhibitor. <u>Exp. Eye Res. 58: 107-116, 1994</u>.
- Pierce, W.M., Jr., M. Sharir, K.J. Waite, D. Chen and K.K. Kaysinger. Topically active ocular carbonic anhydrase inhibitors: Novel (*biscarbonyl*)amidothiadiazoles sulfonamides as ocular hypotensive agents. Proc. Soc. Exp. Biol. Med., <u>203</u>: 360-365, 1993.
- Fish, R.H., K.J. Oberhausen, S. Chen, J.F. Richardson, W.M. Pierce, Jr. and R.M. Buchanan. Biomimetic Oxidation Studies. 7. Alkane functionalization with a MMO structural model [Fe2O(OAc)(tris((1-methylylimidazol-2-yl)methyl)amine)₂]³⁺, in the presence of t-butylhydroperoxide and oxygen gas. Catalysis Letters <u>18</u>: 357-265, 1993.
- Sharir, M., W.M. Pierce, Jr. and T.J. Zimmerman. Topically active ocular carbonic anhydrase inhibitors: Novel (*biscarbonyl*)amidothiadiazoles sulfonamides as probes of corneal endothelial function. J. Ocular Pharmacology 9: 333-340, 1993.
- Lenz, L.G., W.K. Ramp, R.J.S. Galvin and W.M. Pierce, Jr. Inhibition of cell metabolism by a smokeless tobacco extract: Tissue and species specificity. Proc. Soc. Exp. Biol. Med. <u>199</u>: 211-217, 1992.
- Galvin, R.J.S., W.K. Ramp, L.G. Lenz and W.M. Pierce, Jr.. Smokeless tobacco contains an inhibitor of prolyl hydroxylase activity. Toxicol. Lett. <u>62</u>: 301-310, 1992.
- King, K.L., N.A. Delamere, S.C. Csukas and W.M. Pierce, Jr.. Metabolism of arachidonic acid by isolated rabbit ciliary epithelium. Exp. Eye Res. <u>55</u>: 235-241, 1992.
- J.C. Passmore, A.E. Jimenez and W.M. Pierce Jr. Cardiac output and the blood pressure increase in deoxycorticosterone acetate-salt sensitive hypertension after nicotine infusion. Clin. Experimental Hypertension <u>A13</u>:83-102, 1991.
- W.M. Pierce, Jr., G.F. Nardin, M.F. Fuqua, E. Sabah-Maren and S.H. Stern. Effect of chronic carbonic anhydrase inhibitor therapy on bone mineral density in white women. J. Bone and Mineral Res. <u>6</u>: 347-354, 1991.

- Song, W., W.M. Pierce Jr., R.A. Prough, and R.N. Redinger. Characteristics of cholesterol 7- α -hydroxylase and 7- α -hydroxycholesterol hydroxylase activities of rodent liver. Biochem.Pharmacol. 41: 1439-1447, 1991.
- Rodrigues, A.D, D. Fernandez, M.A. Nozarzewski, W.M. Pierce, Jr. and R.A. Prough. Inhibition of hepatic microsomal cytochrome P450-dependent monooxygenase activity by the antioxidant 3-t-butyl-4-hydroxyanisole. Chem. Res. Toxicology 4: 281-289, 1991.
- S.D. Gettings, C.B. Brewer, W.M. Pierce Jr., J.A. Peterson, A.D. Rodrigues and R.A. Prough. Enhanced decomposition of oxyferrous cytochrome P450CIA1 (P-450_{cam}) by the chemopreventive agent, 3-*tert*-butyl-4-hydroxyanisole. Arch. Biochem. Biophys. <u>276</u>: 500-509, 1990.
- T. Yamamoto, W.M. Pierce, Jr., H.E. Hurst, D. Chen and W.J. Waddell. Ethyl carbamate metabolism: <u>in vivo</u> inhibitors and <u>in vitro</u> enzymatic systems. Drug Metabolism and Disposition <u>18</u>: 276-280, 1990.
- F.W. Benz, D.E. Nerland, W.M. Pierce, Jr. and C. Babiuk. Acute acrylonitrile toxicity: Studies on the mechanism of the antidotal effect of D- and L-cysteine and their N-acetyl derivatives. Toxicol. Appl. Pharmacol. <u>102</u>: 142-150, 1990.
- D.E. Nerland and W.M. Pierce, Jr. Identification of N-acetyl-S-(2,5-dihydroxyphenyl)-L-cysteine as a urinary metabolite of benzene, phenol, and hydroquinone. Drug Metabolism and Disposition <u>18</u>: 958-961, 1990.
- K.J. Oberhausen, J.F. Richardson, W.M. Pierce, Jr. and R.M. Buchanan. Synthesis, structure and properties of a N3 tridentate bis-imidazolyl ligand with copper(II). Polyhedron <u>8</u>: 659-668, 1989.
- M.S. Mashuta, W.M. Pierce, Jr. and R.M. Buchanan. Binuclear Schiff base macrocycles. Inorg. Chim. Acta <u>158</u>: 227-237, 1989.
- J.S. Hurst, C.A. Paterson, P. Bhattacherjee and W.M. Pierce, Jr. Effects of ebselen on arachadonate metabolism by ocular and non-ocular tissues. Biochem. Pharmacol. <u>38</u>: 3357-3363, 1989.
- L.A. Carr, P.P. Rowell and W.M. Pierce, Jr. Effects of subchronic nicotine administration on central dopaminergic mechanisms in the rat. Neurochemical Research <u>14</u>: 511-515, 1989.
- W.M. Pierce, Jr., A.O. Clark and H.E. Hurst. Determination of ethyl carbamate by gas chromatography with flame ionization or mass spectrometric detection. J. Official Analytical Chemists. <u>71</u>: 781-784, 1988.
- W.M. Pierce, Jr. and D.E. Nerland. Qualitative and quantitative analyses of phenol, phenylglucuronide and phenylsulfate in urine and plasma by gas chromatography/mass spectrometry. J. Anal Toxicology, 12: 344-347, 1988.
- T. Yamamoto, W.M. Pierce, Jr., H.E. Hurst, D. Chen and W.J. Waddell. Inhibition of the

- metabolism of urethane by ethanol. Drug Metab. Disposition 16: 355-358, 1988.
- M.S. Mashuta, T.N. Doman, W.M. Pierce, Jr. and R.M. Buchanan. Synthesis and characterization of a new binucleating Schiff base macrocycle and its nickel(II) and copper(II) complexes. Inorgan. Chim. Acta <u>145</u>: 21-28, 1988.
- W.J. Waddell, C. Marlowe and W.M. Pierce, Jr. Inhibition of the localization of urethane in mouse tissues by ethanol. Food and Chemical Toxicology <u>25</u>: 527-531, 1987.
- R.D. Gray, W.M. Pierce, Jr., J.W. Harrod, Jr., and J.M. Rademacher. Inhibition of thermolysin by bifunctional N-carboxyalkyl dipeptides. Arch. Biochem. Biophys. <u>256</u>: 692-698, 1987.
- W.M. Pierce, Jr. and L.C. Waite. Bone-targeted carbonic anhydrase inhibitors: Effect of a proinhibitor on bone resorption in vitro. Proc. Soc. Exp. Biol. Med. 186: 96-102, 1987.
- T.I. Senler, W.L. Dean, W.M. Pierce, Jr. and J.L. Witliff. Procedures for measuring Cytochrome P-450-dependent hydroxylation activity in reproductive tissues. Anal. Biochem. 144: 152-158, 1985.
- W.M. Pierce, Jr., J.A. Blank and L.C. Waite. Effects of heterocyclic sulfonamides on bone metabolism. Res. Comm. Chem. Path. Pharm. 50: 3-20, 1985.
- W.M. Pierce, Jr., J.J. Schlager, R.J. Madden and H.E. Hurst. A simple, rapid synthesis of caffeine-1,7-¹³C. J. Labelled Compounds and Radiopharmaceuticals <u>21</u>: 187-192, 1984.
- W.M. Pierce, Jr., M.D. Lineberry and L.C. Waite. Effects of sulfonamides on the hypercalcemic response to Vitamin D. Hormone and Metabolic Res. <u>14</u>: 670-673, 1982.
- W.M. Pierce, Jr. and L.C. Waite. Acetazolamide inhibition of bone resorption: lack of effect on phosphate release from bone <u>in vitro</u>. Hormone and Metabolic Res. <u>13</u>: 591-592, 1981.
- L.C.Waite, W.M. Pierce, Jr. and M.D. Lineberry. Sulfonamide inhibition of bone resorption: lack of a hypophosphatemia. J. Pharmacol. Exp. Ther. 213: 441-444, 1980.

Return to Beginning of Bibliography

BOOK CHAPTERS, REVIEWS, PATENTS

- W.M. Pierce, Jr. and T.J. Zimmerman. The development of topically active ocular carbonic anhydrase inhibitors with potential utility for the management of glaucoma. Transactions of the American Glaucoma Society, 1987.
- L.C. Waite and W.M. Pierce, Jr. Carbonic anhydrase inhibitors and bone metabolism. Proceedings of Biochemical Pharmacology (PRC), Vol. 2, pp. 170-177, December, 1987.
- T.J. Zimmerman and W.M. Pierce, Jr. Book Review: <u>Clinical Ophthalmic Pharmacology.</u> Ed. D.W. Lambert and D.E. Potter. Am. J. Ophthalmol. 104: 558, 1988.
- W.M. Pierce, Jr. Topically active ocular benzothiazole sulfonamide carbonic anhydrase inhibitors. <u>U.S. Patent No. 5,059,613</u>. Issued October 8, 1991.
- W.M. Pierce, Jr. Topically active ocular gem-diacylthiadiazole sulfonamide carbonic anhydrase inhibitors. <u>U.S. Patent No. 5,055,480</u>, Issued October 22, 1991.
- W.M. Pierce, Jr. Topically active ocular thiadiazole sulfonamide carbonic anhydrase inhibitors. <u>U.S. Patent 5,242,937</u>. September 7, 1993.
- W.M. Pierce, Jr. and L.C. Waite. Bone Targeted Carbonic Anhydrase Inhibitors. <u>U.S. Patent 5,641,762</u>. Issued June 24, 1997.

Return to Beginning of Bibliography

ABSTRACTS AND PRESENTATIONS:

- W.M. Pierce, Jr., M.D. Lineberry and L.C. Waite. Inhibition of the hypercalcemia associated with pharmacological doses of Vitamin D3 inhibitors of carbonic anhydrase. Fed. Proc. 38: 849, 1979.
- L.C. Waite, W.M. Pierce, Jr. and M.D. Lineberry. Interrelation of the effects of carbonic anhydrase inhibitors on blood pCO2 and phosphate. The Pharmacologist <u>21</u>: 185, 1979.
- W.M. Pierce, Jr. and L.C. Waite. Acetazolamide inhibits bone resorption <u>in vitro</u> without effect on lactic acid or cAMP production. The Pharmacologist <u>22</u>: 254, 1980.
- W.M. Pierce, Jr., M.D. Lineberry and L.C. Waite. Attenuation of the skeletal response to 1,25-dihydroxyvitamin D3 in vivo and in vitro. Fed. Proc. 39: 729, 1980.
- W.M. Pierce, Jr. and L.C. Waite. Dissociation of PTH-stimulated calcium and phosphate effluxes from bone by acetazolamide. Fed. Proc. 40: 350, 1981
- T.I. Senler, W.L. Dean, W.M. Pierce, Jr. and J.L. Witliff. Cytochrome P-450 dependent hydroxylation in reproductive tissues. Fed. Proc. <u>43</u>: 1475, 1984.
- W.M. Pierce, Jr. and L.C. Waite. Bone targeted inhibitors of carbonic anhydrase. Fed. Proc. <u>45</u>: 1017, 1986.
- W.J. Waddell, C. Marlowe and W.M. Pierce, Jr. Persistent inhibition of urethane uptake in mouse tissues by ethanol. IUPHAR 1987.
- S.D. Gettings, C.B. Brewer, L.S. Crouch, W.M. Pierce, Jr., J.A. Peterson and R.A. Prough. Enhancement of oxy-ferrous P-450_{cam} decomposition in the presence of butylated hydroxyanisole (BHA). Presented at the Annual Meeting of the American Society of Biological Chemists, 1987.
- R.A. Prough, S.D. Gettings, W.M. Pierce, Jr., L.S. Crouch, C.B. Brewer and J.A. Peterson. Interaction of butylated hydroxyanisole (BHA) with oxy- ferrous P-450_{cam} under steady state conditions. 7th International Symposium on Microsomes and Drug Oxidations, Adelaide, South Australia. 1987.
- R.M. Buchanan, M.S. Mashuta and W.M. Pierce Jr. Binucleating N,N' ethylene bis (acetylacetone-iminato) Schiff base macrocycles. 193rd National Meeting of the American Chemical Society, Denver, CO, April 1987 Abs. INORG 140.
- R.M. Buchanan, M.S. Mashuta and W.M. Pierce Jr. Synthesis and characterization of neutral cyclic bis-N2O2 macrocycles. 193rd National Meeting of the American Chemical Society, Denver, CO, April 1987 Abs. INORG 141.
- D.E. Nerland and W.M. Pierce, Jr. Identification of N-acetyl-S-(2,5- dihydroxyphenyl)-L-cysteine in the urine of benzene treated rats. The Pharmacologist, 1987.
- T. Yamamoto, W.M. Pierce, Jr., H.E. Hurst, D. Chen and W.J. Waddell. Inhibition of the metabolism of urethane by ethanol in mice. Proc. ISSX/SOT Symposium on Endogenous Factors in the Toxicity of Xenobiotics. Clearwater, Florida, USA, November, 1987.

- M.K. King and W.M. Pierce, Jr. Monitoring of bone metabolic parameters in vitro. Fed. Proc. 47:1988.
- N. Kurata, R.A. Kemper, H.E. Hurst, W.M. Pierce, Jr. and W.J. Waddell. GC/MS determination of urethane pharmacokinetics in mice. The Toxicologist <u>9:</u>1988.
- D.E. Nerland, F.W. Benz, C. Babiuk and W.M. Pierce, Jr. Effect of cytochrome P-450 inhibitors and ethanol on the acute toxicity of acrylonitrile and its <u>in vivo</u> metabolism to cyanide. The Toxicologist: <u>9</u>, 159 1989.
- A.D. Rodriques, C. Brewer, S.D. Gettings, W.M. Pierce, Jr., J.A. Peterson and R.A. Prough. Uncoupling of cytochrome P-450_{cam} function by 3-t-butyl-4-hydroxyanisole. Fed. Proc. 48:1989.
- D. Chen and W.M. Pierce, Jr. Effects of zinc on bone metabolism in chick tibiae in vitro. The Pharmacologist 1989.
- F.W. Benz, D.E. Nerland, C. Babiuk and W.M. Pierce, Jr. Antidotal effect of the optical isomers of cysteine and N-acetylcysteine on acute acrylonitrile toxicity. The Toxicologist: 9, 284, 1989.
- W. Song, R.A. Prough, W.M. Pierce Jr. and R.N. Redinger. Measurement of 7-alphahydroxycholesterol formed by rodent hepatic microsomes. Presented at the annual meeting of the American Gastroenterological Society, San Antonio, May 1990.
- M. Sharir, K.J. Waite, D. Chen, K.K. Kaysinger and W.M. Pierce, Jr. Novel thiadiazole sulfonamide carbonic anhydrase inhibitors as topically effective ocular hypotensive agents. Invest. Ophthalmol. Vis. Sci. <u>31</u>:149, 1990.
- K. Ochsner, M. Sharir, W.M. Pierce, Jr. and R.A. Eiferman. An in vivo corneal stress test. Invest. Ophthalmol. Vis. Sci. 31:145, 1990.
- W.M. Pierce, Jr., K.K. Kaysinger, L.C. Waite and K.J. Waite. Bone targeted carbonic anhydrase inhibitors: Prodrugs and active configurations. FASEB J.4:, 1990.
- W.M. Pierce, Jr. Aliphatic alcohols and oxidative damage. The Toxicologist 10: 1990.
- K.K. Kaysinger and W.M. Pierce, Jr. Effect of alterations in 2-oxoglutarate metabolism on bone prolyl hydroxylase. FASEB J. <u>4</u>: 1990.
- A.D. Rodrigues, D. Fernandez, W.M. Pierce, Jr., J.A. Peterson and R.A. Prough. The role of 3-t-butyl-4-hydroxyanisole in inhibition of cytochrome P-450 function. Antioxidant Symposium, Berkeley, CA, January, 1990.
- D. Chen and W.M. Pierce, Jr. Effects of Zinc and PGE2 on in vitro Bone Metabolism. FASEB J. <u>5</u>:1991.
- R.J.S. Galvin, L.G. Lenz, W.K. Ramp and W.M. Pierce, Jr. Smokeless tobacco contains a competitive inhibitor of prolyl hydroxylase. First International Conference on Smokeless

- Tobacco and Health, Columbus, OH, April 1991.
- W.K. Ramp, L.G. Lenz, K.K. Kaysinger and W.M. Pierce, Jr. Interrelationships of parathyroid hormone, collagen synthesis and alkaline phosphatase in osteoblast-like cells. XIth International Congress of Calcium Regulating Hormones, Florence, Italy, April, 1992.
- R.M. Buchanan, S. Chen, J. Wang, M.N. Balachander, H. Nie, J.F. Richardson, M.S. Mashuta, N. Smith, W.M. Pierce, Jr. and R.H. Fish. Metal Catalyzed Functionalization of C-H bonds. A Structure-Activity Study. 205th National Meeting of the American Chemical Society, Washington, DC, 1994.
- W. Song, W.M. Pierce, Jr., Y. Saeki, R.N. Redinger and R.A. Prough. 7α Hydroxycholesterol Oxidoreductase: A Novel Enzyme found in Hamster and Human Liver. Annual Meeting of the American Society for Biochemistry and Molecular Biology, 1994.
- G.C. Rodgers, Jr., W.M. Pierce, Jr., F. Jordachescu, S. Fpiru, M. Nanulescu, C. Pop and D. Mararu. Lead Poisoning in Romania. Child Health 2000, World Congress and Exposition on Child Health, Vancouver, Canada, June 1995.
- Wu, Q., N.A. Delamere and W.M. Pierce, Jr. Carbonic anhydrase isozymes in non-pigmented ciliary epithelial cells. Association for Research in Vision and Ophthalmology, Annual Meeting, Fort Lauderdale, FL, April 21-26, 1996. INVEST OPHTH VIS SCI 37: (3) 5100-5100 FEB 15 1996.
- Pierce, W.M., Jr., D. Chen and L.C. Waite. Zinc stimulates osteoblastic collagen synthesis. American Society of Bone and Mineral Research, Annual Meeting, Seattle, WA, September, 1996.
- Pierce, W.M Jr., D.E. Rueff, N.B. Smith and G. C. Rodgers, Jr. Blood Lead Concentrations in the Children of Jefferson, County, Kentucky. Presented at the Annual Meeting of the Kentucky Academy of Sciences, November, 1997.
- Hou Y, Pierce WM, Delamere NA Ascorbic acid alters sodium transport in cultured rabbit ciliary epithelium INVEST OPHTH VIS SCI 38: (4) 4864-4864 Part 2 MAR 15 1997
- Spatola, A.F., P.J. Romanovskis and W.M. Pierce, Jr. Cyclic Pseudopeptides. Presented at International Peptide Symposium, Vienna, Austria, Fall, 1998.
- Jason R. Neale, Nicole K. Emery, Ned B. Smith, Sujan Singh, Leonard C. Waite, K. Grant Taylor and William M. Pierce, Jr.A Bone Targeting Strategy for Selective Treatment of Bone Metabolic Disorders. Presented at the Annual Meeting of the American Society for Bone and Mineral Research, St. Louis, MO, USA, October, 1999.
- Li J, Benz FW, Pierce WM, Feldhoff RC, and Nerland DE. Identification of the predominant site of covalent binding of acrylonitrile to rat hemoglobin. Toxicological Sciences 48: No1-Supplement, 158, 1999.
- L. R. Shoemaker, J.J. Miller, W.M. Pierce, Jr., S.J. Schurman, W.H. Bergstrom.

Furosemide-induced hypercalciuria: A model of hypercalciuric Bartter syndrome (HBS) Pediatric Res, 2000.

William M. Pierce Jr., K. Grant Taylor, Leonard C. Waite, Sujan Singh, Jason R. Neale, Xiaoping Tang, and Ned B. Smith. Bone-targeted Estrogens: Anabolic Bone Effects of an Ether-linked 17-O-Estradiol Derivative. J. Bone Mineral Res., 2000 Presented at the Annual Meeting of the American Society for Bone and Mineral Research.

K. Grant Taylor, Jason R. Neale, Sujan Singh, Xiaoping Tang, Peter C. Kulakosky, Valentyn V. Tyulmenkov, Leonard C. Waite, Carolyn M. Klinge, and William M. Pierce, Jr. BONE SELECTIVE ESTROGENS: Estrogen Receptor Alpha Selectivity is a Predictor of in vivo Efficacy. Presented at the annual meeting of the American Chemical Society - Southeast / Southwest Regions.

E. Gozal, J.B. Klein, W.M. Pierce, J.A. Scherzer. J.Cai, L.R. Sachleben and D. Gozal. Proteomic analysius of CA1 and CA3 regions of the hippocampus following 6 hours of intermittent hypoxia. Presented at the annual meeting of the Society for Neurosciences, 2000.

Klein, J.B., J.J. Williams, J.A. Scherzer, J. Cai, W.M. Pierce and J.M. Arthur. Development of a protein expression database and comparison of rat renal cortical and medullary protein expression using high-throughput proteomic analysis. . J. Am. Soc. Nephrol. 11:409A, 2000.

Arthur, J.M., J.J. Williams, J.A. Scherzer, J. Cai, W.M. Pierce and J.B. Klein. Proteomic identification of proteins involved in magnesium reabsorption in the kidney. J. Am. Soc. Nephrol. 11:557A, 2000.

Parathyroid hormone (PTH) regulates sodium phosphate cotransport (NaPi-4) through an A kinase anchoring protein (AKAP) Khundmiri SJ, Klein JB, Pierce WM, Lederer ED FASEB Journal 15 (4): A144, Part 1 MAR 7 2001

CV Rao, JR Neale, S Mishra, WM Pierce and ZM Lei. Bone changes in LH receptor knockout animals. To be presented at the Annual Meeting of the Endocrine Society, Denver, CO June, 2001.

Perez-Abadia, G, VS Gorantla, M Vossen, MJ Voor, CG Francois, X Ren, PCR Brouha, H Orhun, R Majzoub, KA Prabhune, WM Pierce, C Maldonado and JH Barker. Assessing bone allograft rejection by measuring mechnaical properties of bone. To be presented at the Plastic Surgery Research Council National Meeting, Milwaukee, WI, June, 2001.

M Vossen, G Perez-Abadia, VS Gorantla, MJ Voor, CG Francois, X Ren, PCR Brouha, H Orhun, R Majzoub, KA Prabhune, WM Pierce, M Kon, C Maldonado and JH Barker. Effects of mixed allogeneic chimerism on the mechanical properties of bone. To be presented at the Plastic Surgery Research Council National Meeting, Milwaukee, WI, June, 2001.

Gozal E, Klein JB, Pierce WM, Scherzer JA, Sachleben Jr. LR, Cai J, Gozal D.

Intermittent hypoxia elicits differential responses in the CA1 and CA3 regions of the rat hippocampus: a proteomic analysis. Presented at: 15th Annual Associated Professional Sleep Societies Meeting, June 5-10, 2001, Chicago, IL Abstracted in: Sleep 2001; 24:A264

PLASMA MEMBRANE Ca2+-ATPase IS PHOSPHORYLATED ON TYROSINE 1176 IN ACTIVATED HUMAN PLATELETS. William L. Dean1, Tina C. Wan1, Jian Cai2 and William M. Pierce, Jr.2

Return to Beginning of Bibliography

CH₃CH₂OCH₂CH₃ Ethoxyethane (Diethyl ether)



(1,4-Dioxane)

CH₃OCH₂CH₂OCH₃
1,2-Dimethoxyethane
(Glycol dimethyl ether,

glyme)



Oxacyclopentaine (Tetrahydrofuran, THF)

Cyclic ethers are members of a class of cycloalkanes in which one or more carbons have been replaced by a *heteroatom*—in this case, oxygen. (A **heteroatom** is defined as any atom except carbon and hydrogen.) Cyclic compounds of this type, called **heterocycles**, are discussed more fully in Chapter 25.

The simplest system for naming cyclic ethers is based on the **oxacycloalkane** stem, in which the prefix *oxa* indicates the replacement of carbon by oxygen in the ring. Thus, three-membered cyclic ethers are oxacyclopropanes (other names used are oxiranes, epoxides, and ethylene oxides), four-membered systems are oxacyclobutanes, and the next two higher homologs are oxacyclopentanes (tetrahydrofurans) and oxacyclohexanes (tetrahydropyrans). The compounds are numbered by starting at the oxygen and proceeding around the ring.

The physical properties of ethers reflect the absence of hydrogen bonding

The molecular formula of simple alkoxyalkanes is $C_nH_{2n+2}O$, identical with that of the alkanols. However, because of the absence of hydrogen bonding, the boiling points of ethers are much lower than those of the corresponding isomeric alcohols (Table 9-1). The two smallest members of the series are water miscible, but ethers become less water soluble as the hydrocarbon residues increase in size. For example, methoxymethane is completely water soluble, whereas ethoxyethane forms only an approximately 10% aqueous solution.

In summary, ethers can be named as alkoxyalkanes or as dialkyl ethers. They have lower boiling points than comparable alcohols because they cannot enter into hydrogen bonding with themselves.

TABLE 9-1 Boiling Points of Ethers and the Isomeric 1-Alkanols					
Ether	Name	Boiling point (°C)	1-Alkanol	Boiling point (°C)	
CH ₃ OCH ₃	Methoxymethane (Dimethyl ether)	-23	CH ₃ CH ₂ OH	78.5	
CH ₃ OCH ₂ CH ₃	Methoxyethane (Ethyl methyl ether)	10.8	CH ₃ CH ₂ CH ₂ OH	82.4	
CH ₃ CH ₂ OCH ₂ CH ₃	Ethoxyethane (Diethyl ether)	34.5	CH ₃ (CH ₂) ₃ OH	117.3	
(CH ₃ CH ₂ CH ₂ CH ₂) ₂ O	1-Butoxybûtane (Dibutyl ether)	142	CH ₃ (CH ₂) ₇ OH	194.5	

5 291



10

15

20

25

Substitute Specification

Version with Markings to Show Changes Made

Attorney Docket: 1540/139

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Alkyl Ether Modified Polycyclic Compounds Having a Terminal Phenol and Uses for Protection of Cells

Inventors: Laszlo Prokai and James W. Simpkins

Government Rights

This patent was created with support from the National Institute on Aging under grant number POI 10485. The US Government has certain rights to the invention.

Cross Reference to Related Applications

This application gains priority from the provisional application filed June 27, 2000 herein incorporated by reference.

Technical Field and Background Art

The present invention relates to methods and compositions to achieve a cytoprotective effect concerning a polycyclic compound with a phenol group at a first end and a carbon ring at a second end in which the hydroxy group on the carbon ring has been substituted by an alkyl ether group.

The naturally occurring hormone 17β-estradiol plays a pivotal role in sexual reproduction in humans and other mammals. It is believed that this estrogenic activity is orchestrated through the binding of estrogen receptors on the surface of target cells (Gridley et al. (1998) Vol. 54, pp. 874-880). Estrogen compounds including 17β-estradiol have also been shown to have neuroprotective activity (US 5,554,601). More generally, cytoprotective activity has been demonstrated for estrogen compounds that have little or no estrogenic activity and in addition have low or [negligable] negligible binding affinity for the estrogen receptor (US 5,843,934). An important functional group in these molecules that determine cytoprotection is the presence of a terminal phenolic group. This observation led to the realization that polycyclic compounds had neuroprotective activity contingent on the presence of a terminal phenol group. (US 5,859,001, 6,197,833) (Bishop et al. (1994) Mol. Cell. Neurosci, Vol. 5, pp. 303-308; Green et al. (1997) J. Steroid Biochem. Mol. Biol., Vol. 63, pp. 229-235).

The above described cytoprotective activity has numerous uses in protecting cells in vivo and in vitro from degeneration that may occur through disease, trauma or aging.

Treatment based on cytoprotection can lead to the slowing of progression of degeneration and postpone the onset of symptoms associated with degeneration. It is desirable therefore, to identify improvements in cytoprotective compounds that might enhance their bioactivity.

Summary of the Invention

5

10

15

20

25

30

A first embodiment of the invention provides a cytoprotective compound that includes a polycyclic compound optionally having two, three or four carbon rings, the compound also having a first end and a second end wherein a phenol group is located at the first end and a terminal carbon ring is located at the second end, the terminal carbon ring having an alkyl ether functional group, the alkyl portion of which having a formula $[C_n H_{2n+2}]$ $C_n H_{2n+2}$ wherein n is at least 3 and less than 20.

In additional embodiments, the carbon ring at the second end is a D ring in a four ring compound which may be an estrogen. The four ring estrogen compound may include an alkyl ether group in an alpha or beta orientation. Moreover the alkyl ether functional group can include any of a long chain saturated alkyl, a long chain unsaturated alkyl, or a cycloalkyl group. In specific embodiments, the cytoprotective compound may be a 17-butoxyestra 1,3,5(10) triene-3-ol, 17-pentoxyestra 1, 3, 5 (10) triene-3-ol a 17-hexoxyestra 1,3,5(10) triene-3-ol, a 17 septoxyestra 1,3,5(10) triene-3-ol, or a 17-octyloxyestra 1,3,5(10) triene-3-ol.

In a second embodiment of the invention, the cytoprotective compound includes an estrogen compound having a terminal phenol group at a first end of the compound and a carbon ring at a second end of the compound, the carbon ring at the second end having an alkyl ether functional group wherein the alkyl group has a formula $[C_n H_{2n+2}]$ $C_n H_{2n+2}$ wherein n is at least 3 and less than 20.

In a third embodiment of the invention, a pharmaceutical formulation is provided that includes a cytoprotection effective dose of a polycyclic compound having a phenolic ring at a first terminal position, optionally any of one, two or three additional ring structures and an alkyl ether functional group on a carbon ring in a second terminal position.

In a fourth embodiment of the invention, a method is provided for retarding the development of a degenerative condition associated with a population of cells in a subject, that includes administering to the subject predisposed to the degenerative condition, an effective amount of a polycyclic phenolic compound in a physiologically acceptable formulation, the polycyclic phenolic compound having a phenol located at a first terminal

position, optionally any of one, two or three additional ring structures; the compound having an alkyl ester located on a carbon ring at a second terminal position, the compound retarding the development of the degenerative condition. The method may utilize any of the alkyl ether compounds described herein including four ring compounds with an alkyl ether on carbon 17 of the D ring in an alpha or beta orientation and may further include enantiomers, [diastomers] diastereomers, salts, derivatives and analogs.

5

10

15

20

25

30

The population of cells or tissues may be selected from stem cells, blood cells, epithelial cells, stromal cells including connective tissue cells, neuronal cells, muscle tissue cells, endocrine tissue cells, whole organ cells, bone cells, eye cells, skin cells, reproductive tract cells and urinary tract cells. The degenerative condition may include cardiac, eye, bone, neurodegenerative or ischemic degeneration.

In a fifth embodiment of the invention, a method is provided for synthesizing an estrogen compound having a phenolic A ring and an alkyl ether functional group on carbon 17, that includes: protecting -OH on the phenolic A ring; alkylating the 17-OH with an alkylating agent in the presence of a strong base; removing the protecting group from [-OH] -OH on the phenolic A ring; and purifying the 17- alkyl ether estrogen compound. Moreover, the [-OH] -OH may be on the carbon 3-position and the 17-OH may be in an alpha or beta position. The alkylating agent may be selected from a group consisting of an alkyl halide, a dialkyl sulfate and an alkyl tosylate. The phenolic-OH may be treated with a base resistant protecting group such as tert-butyl, methoxymethyl and 9-anthrylmethyl. The protecting group may be removable by acid hydrolysis, catalytic hydrogenolysis where the hydrogenolysis may include CF₃COOH or by catalytic transfer hydrogenation which may use ammonium formate. The strong base of the method may include sodium hydride.

In a sixth embodiment of the invention, a method is provided for treating a subject having a degenerative disorder, comprising: obtaining at least one 17-O-alkyl ether of estrogen in a pharmaceutical formulation; and administering an effective dose of the 17-O-alkyl ether of estrogen to the subject so as to treat the degenerative disorder.

In a seventh embodiment of the invention, a method is provided for conferring cytoprotection of a population of cells, that includes providing an 17β-O-alkyl ether of an estrogen compound; and administering the compound in an effective dose to the population of cells so as to confer cytoprotection on the population of cells. All embodiments directed to methods include the use of any of the alkyl ether compounds described herein

Brief Description of the Drawings

The foregoing features of the invention will be more readily understood by reference to the following detailed description, taken with reference to the accompanying drawings, in which:

Figure 1 shows the structure of the alkyl ether of estradiol.

5

10

15

20

25

30

Figure 2 shows the synthesis of 17-alkyl ether of estradiol.

Figure 3 is an ORTEP plot of the X-ray crystal structure of 17-O-butylated 17 β -estradiol (4d). Thermal ellipsoids are shown at the 30% probability level.

Figure 4 shows a graphical representation of cell viability, where the cells are HT-22 cell cultures after glutamate exposure (20 mM) (a) following treatment with estradiol and its 17β -alkyl ethers (4a-4f), and 3-butyl estradiol (5b as a typical representative of the 3-alkyl ethers). Statistically significant differences between groups were tested by analysis of variance (ANOVA) followed by post hoc Tukey test: * significant increase (p <0.05) vs vehicle control, ** significant increase (p <0.05) vs vehicle control, but decrease compared to 10μ M estradiol (1), *** increase (p <0.05) vs vehicle control, and statistically significant increase compared to 10μ M estradiol (1).

Detailed Description of Specific Embodiments

As used in this description and the accompanying claims, the following terms shall have the meanings indicated, unless the context otherwise requires:

"Estrogen compound" is defined here and in the claims as any of the structures described in the 11th edition of "Steroids" from Steraloids, Inc. Wilton, NH, here incorporated by reference. Included in this definition are non-steroidal estrogens described in the aforementioned reference. Other estrogen compounds included in this definition are [eyclopenantophenanthrene] cyclopentanophenanthrene compounds, estrogen derivatives, estrogen metabolites and estrogen precursors as well as those molecules capable of binding cell associated estrogen receptor as well as other molecules where the result of binding specifically triggers a characterized estrogen effect. Assumed as included in this definition but more explicitly stated, are isomers, [diasteromers] diastereomers and enantiomers of the aforementioned, as well as mixtures of more than one estrogen.

In an embodiment of the invention, "cytoprotective effect" is a measurable positive effect on the survival of cells that would otherwise die without an intervention.

"Treatment" of a disorder in a patient with a cytoprotective compound may be characterized as, but is not limited to, a slowing of progression of a disorder and optionally "Treatment" of a disorder in a patient with a cytoprotective compound may be characterized as, but is not limited to, a slowing of progression of a disorder and optionally slowing of the development of symptoms than would otherwise occur in the absence of the compound.

"Alkyl ether functional group on the carbon ring at the second end" includes locating the alkyl ether functional group on any available carbon in the ring for example, carbon-15,
16 or -17.

"terminal Terminal phenol group" includes a carbon ring with an OH- group on any of carbons 2, 3 or 4.

"Alkyl ether functional group on carbon 17 of the D ring "refers unless specified otherwise to 17 β -, 17 α -, enantiomers of the four ring compound, salts, derivatives and analogs thereof. Similarly, a 17-alkylestra-1, 3,5(10) triene-3-ol refers to any of the 17- α or 17- β [diasteromers] diastereomers, and the enantiomers of the compound, salts, derivatives and analogs thereof.

"17-" refers to 17 β - or 17 α -.

5

10

15

20

25

30

We have synthesized novel modifications of known compounds that have improved cytoprotective activity when compared with the unmodified forms. The novel compounds are polycyclic compounds with a terminal phenol group that have been modified in such a way as to increase the lipophilicity of the compounds for improved uptake by target cells thereby improving the cytoprotective effect of the compounds while maintaining the terminal phenol group. Polycyclic compounds with a terminal phenol group prior to modification with an alkyl ether as described below include those compounds listed in US patent 6,197,833 herein incorporated by reference. Embodiments of the invention include compounds with significantly less feminizing activity compared with 17β-estradiol and include compounds that do not readily bind the estrogen receptor (Table 10). Accordingly, modifications include the addition of an alkyl ether on carbon 17 of the molecule, where the alkyl group is characterized by the formula C_nH_{2n+1} in which n is at least 3 and less than 20 more particularly, where n=3-16, more particularly where n=3-12, more particularly where n=3-8. A limitation on the length of the alkyl ether resides in the solubility of the compound in solvents suitable for delivery of the compound to a subject by an appropriate route of delivery selected to achieve either acute or chronic administration. Examples of solvents are provided below. The alkyl ether modification may further include cyclical alkyl ethers including cyclohexyl and cyclopentyl derivatives.

group consisting of an alkyl halide, a dialkyl sulfate and an alkyl tosylate. The phenolic-OH may be treated with a base resistant protecting group such as tert-butyl, methoxymethyl and 9-anthrylmethyl. The protecting group may be removed by acid hydrolysis, catalytic hydrogenolysis where the hydrogenolysis may include CF₃COOH or by catalytic transfer hydrogenation which may use ammonium formate. The strong base of the method may include sodium hydride.

In an embodiment of the invention, an alkyl ether substituted 17-β estradiol is shown schematically in Figure 1. In addition, the synthetic pathway for making 17-alkyl ether of estradiol is shown in Figure 2 with a crystallographic structure of 17-O-butylated 17-β estradiol in Figure 3. The cytoprotection provided by alkyl ether compounds as described has been demonstrated in HT22 assays. (Figure 4) The observed cytoprotective effect is independent of estrogenic normal activity. Cytoprotective activity using these compounds is not limited to HT22 cells but is applicable to different cell populations and tissues found in a subject and present *in vivo* and *in vitro* regardless of whether those cells carried an estrogen receptor or not.

10

15

20

25

30

The experimental models for measuring cytoprotection have become established using a range of cell cultures such as HT22, (described below in the Example 2) SK-N-SH (American Type Culture Collection, Rockville, MD) described in US Patent 5,554,601, erythrocytes and muscle cells and in *in vivo* animal models. Experimental animals such as rats have been described in which a traumatic event such as ovariectomy itself or additional insult such as an arterial occlusion is generated in ovariectomized and non-ovariectomized animals. (US Patents 5,554,601, and 5,859,001). The treated and non-treated rats are then measured for the cytoprotective effect afforded by a range of doses of the compound administered to the animal subject.

The cytoprotective compounds described herein can be used in effective doses to treat patients with acute or chronic degenerative disorders. Examples of acute degenerative disorders include: tissue ischemic events (US patent 5,877,169, herein incorporated by reference), for example, cerebrovascular disease, subarachnoid hemorrhage or trauma, prevention of ischemia reperfusion injury, prevention of ischemia reperfusion injury in the setting of resuscitation from hypovolemic shock, renal ischemia, myocardial infarction, angina and cardiac ischemia, endothelial inflammation, and cardiotoxicity associated with administration of anti-cancer compositions. Similarly, effective doses of the cytoprotective compounds may be beneficial in treating osteoporosis. (US Patent 5,843,934 herein incorporated by reference). Moreover, the compounds may be used to protect cells in graft

tissue during transplantation. (US Patents 5,824,672 and 6,207,658 herein incorporated by reference) The compounds may be used to protect aging skin and skin damaged by cytotoxic events either in a cosmetic formulation or as a therapeutic agent. The compounds may be used to protect against vascular degeneration associated with diabetes.

Graft cells include those cells, tissues or organs obtained from a donor by transplantation into a recipient, where the graft cells may be derived from human subjects or from animals and may be transplanted from one subject back into the same subject or from one subject (the donor) into another subject (the recipient) for improving the health of the recipient. In these situations, the donor subject can be a living subject, fetus or a recently deceased subject. The grafts cells and tissues include stem cells, blood cells, bone marrow cells, placental cells, sperm and ova and may further include heart, lungs, corneal tissue or fetal tissue. Accordingly, the compounds described herein may be beneficial in protecting graft cells from damage resulting from oxidative stress.

The cytoprotective compounds described herein can be used to protect neurons from severe degeneration and is an important aspect of treatment for patients with acute or chronic neurodegenerative disorders. Examples of chronic disease include Alzheimer's disease. (US 5,554,601 herein incorporated by reference), Parkinson's disease, Huntingdon's disease, AIDS dementia, Wernicke-Korsakoff's related dementia (alcohol induced dementia), age related dementia, age associated memory impairment, brain cell loss due to any of the following: head trauma, stroke, myocardial infarction, hypoglycemia, ischemia, anoxia, [hypoxia] hypoxia, cerebral edema, arteriosclerosis, diabetic neuropathy, hematoma and epilepsy, spinal cord cell loss due to any of the conditions listed under brain cell loss; and peripheral neuropathy.

Other examples of degenerative diseases, disorders and conditions that may be treatable by a cytoprotective agent include: various bone disorders including osteoporosis, osteomyelitis, ischemic bone disease, fibrous dysplasia, rickets, Cushing's syndrome and osteoarthritis, other types of arthritis and conditions of connective tissue and cartilage degeneration including rheumatoid, psoriatic and infectious arthritis, various infectious diseases, muscle wasting disorders such as muscular dystrophy, skin disorders such as dermatitis, eczema, psoriasis and skin aging, degenerative disorders of the eye including macular degeneration and retinal degeneration, disorder of the ear such as otosclerosis, impaired wound healing, various diseases and conditions of the heart including cardiac ischemia, myocardial infarction, chronic or acute heart failure, cardiac [dysrhymias, artrial] dysrhythmias, arterial fibrillation, [paroxymial] paroxymal tachycardia, ventricular

fibrillation and congestive heart failure, circulatory disorders including atherosclerosis, arterial sclerosis and peripheral vascular disease, diabetes (Type I or Type II), various diseases of the lung including pneumonia, chronic obstructive lung disease (bronchitis, [emphysemia] emphysema, asthma), disorders of the gastrointestinal tract such as ulcers and hernia, dental conditions such as periodontitis, liver diseases including hepatitis and cirrhosis, pancreatic ailments including acute pancreatitis, kidney diseases such as acute renal failure and [glomerulonepritis] glomerulonephritis, various blood disorders such as vascular amyloidosis, [anerysms] aneurysms, anemia, [hemorrage] hemorrhage, sickle cell anemia, autoimmune disease, red blood cell fragmentation syndrome, neutropenia, leukopenia, bone marrow aphasia, pancytopenia, thrombocytopenia, hemophilia. The preceding list of diseases and conditions which are potentially treatable with a cytoprotective agent is not intended to be exhaustive or limiting but presented as examples of such degenerative diseases and conditions.

10

15

20

25

30

The present compositions may be used for protecting cells including any of the below listed cells or tissues and for treatment of disorders including any of the aforementioned degenerative conditions. Examples of cells that may be protected by the compounds include: stem cells, blood cells, epithelial cells, stromal cells including connective tissue cells, neuronal cells, muscle tissue cells, endocrine tissue cells, whole organ cells, bone cells, skin cells, eye cells, reproductive tract cells and urinary tract cells and tissues that include more than one cell type. Tissues that are protected by the method of the invention may be derived from children, adult or fetal tissue and include, but are not limited to blood and all of its components, including erythrocytes, leukocytes, platelets, serum, central nervous tissue, including brain and spinal cord tissue, neurons, and glia; peripheral nervous tissue, including ganglia, posterior pituitary gland, adrenal medulla, and pineal; connective tissue, including skin, ligaments, tendons, and fibroblasts; muscle tissue, including skeletal, smooth and cardiac tissues or the cells therefrom; endocrine tissue, including anterior pituitary gland, thyroid gland, parathyroid gland, adrenal cortex, pancreas and its subparts, testes, ovaries, placenta, and the endocrine cells that are a part of each of these tissues; blood vessels, including arteries, veins, capillaries and the cells from these vessels; lung tissue; heart tissue and whole organ; heart valves; liver; kidney; intestines; bone, including osteocytes, osteoblasts and osteoclasts; immune tissue, including blood cells, bone marrow and spleen; eyes and their parts; reproductive tract tissues; or urinary tract tissue.

The present compounds may be administered to a subject orally, topically, transdermally through skin or via the mucosal membrane for example the nasal mucosa and

buccal mucosa, or parenterally including intravenous, intramuscular and subcutaneous administration. The compound may be further administered subcutaneously using an oil delivery vehicle for improved uptake and sustained effectiveness. Depending on the intended mode, the compositions may be in the form of solid, semi-solid or liquid dosage forms such as for example, tablets, suppositories, pills, capsules, powders, liquids, suspensions, patches, creams, gels, or the like preferably in unit dosage forms suitable for single administration of precise dosages.

5

10

15

20

25

30

The present compositions can be formulated using suitable solvents including cyclodextrin, various proteins, oils such as, corn oil or sesame oil, or alcohols, the solvents of choice being dependent on the route of administration and the need for sustained delivery. For example, intravenous administration of the composition would utilize an aqueous solvent, whereas subcutaneous delivery of the composition might utilize an oil solvent. The therapeutic formulations will include a conventional pharmaceutical carrier or excipient and a therapeutically effective amount of the active agent (cytoprotective compound) and in addition, may include for example, other therapeutic agents, carriers, adjuvants.

The amount of active compound administered will depend on the human or animal subject being treated, the severity of the condition, the manner of administration and the judgement of the prescribing clinician.

Typical compositions contain approximately 0.01-95% by weight of active ingredient with the balance one or more acceptable non-toxic carriers. The percentage of active ingredient will depend upon the dosage form and the mode of administration. Standard formulations have been enumerated in US Patent 6,020,510 (incorporated by reference) and are similarly applicable herein. An effective dose of the active agent as measured in the plasma of a subject may be for example in the range of 5pg/ml-5000pg/ml.

All references recited herein are incorporated by reference. The following examples are presented to further illustrate embodiments of the invention but are not intended to be limiting.

Examples

Example 1: Method of synthesis of a 17-alkyl ether of 17β-estradiol.

We selectively (and reversibly) protected the 3-OH before alkylating on the 17 position of 17β-estradiol under strong basic condition with the relevant alkyl halide. because alkylation on the phenolic 3-hydroxyl group proceeds under much milder condition than that of the 17 position. Protection of the 3-OH of 17β-estradiol (1) as benzyl (Bz) ether (2) (Qian et al. (1988) J. Steroid Biochem, Vol. 29, pp. 657-664) was achieved

condition than that of the 17 position. Protection of the 3-OH of 17β-estradiol (1) as benzyl (Bz) ether (2) (Qian et al. (1988) J. Steroid Biochem, Vol. 29, pp. 657-664) was achieved by elaboration of the 17β-OH to the corresponding 17β-alkoxyl congeners (3a-f). The 17β-OH group was successfully alkylated with the corresponding alkyl halide in the presence of sodium hydride in dimethylformamide. The 3-benzyl protecting group was removed rapidly under ambient conditions by catalytic transfer hydrogenation using ammonium formate resulting in the desired products (4a-f). (Anwer, et al. (1980) Synthesis, pp. 929-932; Elamin, et al. (1979) J. Org. Chem., Vol. 44, pp. 3442-3444). 3-O-Butyl and octyl ethers of 1(5b,c; Scheme 1) as controls were prepared directly from (1) by using alkyl halide in the presence of potassium carbonate. (The number numbers in parenthesis refer to those in Figure 2.)

5

10

20

25

30

In addition to NMR, mass spectrometry, chromatographic and combustion analyses to characterize the compounds prepared, crystallography data were obtained for two representative 17β -ethers (methoxy and butoxy groups). Summary data for 4d is provided in Table I. The solid-state conformation (ORTEP-type plot) of 4d is shown in Figure 3. The crystals were monoclinic and belonged to the P2 (1) space group, and confirmed that the 17-methoxy and butoxy groups assumed β -orientation in the D-ring.

Instruments and Materials. All solvents and material were obtained from FisherScientific (Atlanta, GA) or from Aldrich (Milwaukee, WI). Estradiol (1) and 3-O methyl-17β-estradiol (5a) were purchased from Sigma (St. Louis, MO). Sodium hydride was used as a 60% dispersion in mineral oil. Melting points were determined on a Fisher-Johns melting point apparatus. Thin layer chromatography (TLC) was done on Whatman silica gel plates (on aluminum backing) containing UV fluorescence indicator. All chromatographic purifications were done on gravity columns with 230-435 mesh neutral silica gel using ethyl acetate: hexane 1:4 (v/v) eluent. Elemental analyses were performed by the Atlantic Microlab, Inc. (Norcross, GA). NMR spectral data were recorded for all compounds using a Varian XL-300 spectrometer using TMS as internal standard. Mass spectral data were obtained by using atmospheric-pressure chemical ionization (APCI) on a quadrupole ion trap instrument (LCQ, Finnigan MAT, San Jose, CA). Analytical reversed-phase high-performance liquid chromatography was performed on a Thermo Separation/SpectraPhysics (Fremont, CA) system consisting of an SP8810 isocratic pump, a Rheodyne (Cotati, CA) Model 7125 injector valve equipped with a 20-µl sample loop, an SP8450 variable wavelength UV/VIS detector operated at 280 nm, and an SP4290 computing integrator. A 15cm x 4.6 mm id. octadecylsilica column (Phase Sep S5 ODS2,

Queensferry, Clwyd, UK) and a mobile phase of acetonitrile containing 1% acetic acid at a flow rate of 1.0 mL/min were used for the analyses.

5

10

15

20

25

30

X-ray crystallography data were collected at 173 K on a Siemens SMART PLATFORM equipped with A CCD area detector and a graphite monochromator utilizing MoK α radiation (1=0.71073 Å). Cell parameters for each structure were refined using up to 8192 reflections and a hemisphere of data (1381 frames) was collected using the w-scan method (0.3° frame width). The first 50 frames were remeasured at the end of data collection to monitor instrument and crystal stability (maximum correction on I was < 1 %). Absorption corrections by integration were applied based on measured indexed crystal faces. Both structures were solved by the Direct Methods in SHELXTL5, (Sheldrick, G. M. (1998). SHELXTL5. Bruker-AXS, Madison, Wisconsin, USA) and refined using full-matrix least squares. The non-H atoms were treated anisotropically, whereas the hydrogen atoms were calculated in ideal positions and were riding on their respective carbon atoms, except the hydroxyl protons H₁₈ in 4a and H₁₈ and H₂₆ in 4d. These protons were obtained from a Difference Fourier map and refined without any constraints. While no solvent crystallized with 4a, a methanol molecule was found in general position in the lattice of 4d. A total of 196 parameters of 4a were refined in the final cycle of refinement using 2961 reflections with I > $2\sigma(I)$ to yield R₁ and wR₂ of 5.03% and 12.66%, respectively. For 4d, a total of 247 parameters were refined in the final cycle of refinement using 3294 reflections with $I > 2\sigma(I)$ to yield R_1 and wR_2 of 3.71% and 8.90%, respectively. Refinement was done using F^2 . Tables of geometric data, indicating H-bonding interactions are provided here for one compound and are further available on the Cambridge Data base for crystallography. (Steps in the synthetic pathway are shown in Figure 2.)

3-Benzyloxyestra-1, 3,5(10)-trien-173-o1 (2). (Quian et al. (1988) J. Steroid Biochem, Vol. 29, pp. 657-664). Benzyl bromide was added to 5 g (18 mmol) of 1 and 10 g (72 mmol) potassium carbonate in 100 ml of acetone 5.7g (4.0mL, 34 mmol). The mixture was refluxed overnight. Upon cooling the solid was removed by filtration. The filtrate was collected and acetone was removed *in vacuo* leaving behind clear yellowish oil, which solidified on standing. Recrystallization from ethyl acetate/hexane gave 6.1g (93% yield) of a white fluffy solid, m.p. 119-121°C; TLC R_f 0.23; ¹H-NMR (CDC1₃) δ: 7.44-7.19 (m, 5H); 6.78(dd, J=8.7 Hz and J=2.7 Hz, 1H); 6.72 (d, J=2.4 Hz, IH); 5.05 (s, 3H); 3.37 (tr, J=8.4 HZ, 1H); 2.87-2.82 (m, 2H); 2.34-1.18 (m, H); 0.78 (s, 3H). MS: *m/z* 363 [M+H]⁺. General Procedure for the Preparation of 3-Benzyloxy-17β-alkoxyestra-1, 3,5(10)-triene (3a-f). Compound 2 (2) (0.8 g, 2.2 mmol) was dissolved in 5 ml anhydrous DMF and, then,

- reaction mixture was quenched by pouring it into 20 mL of dilute hydrochloric acid and extracted with methylene chloride. The organic phase was dried over Na₂SO₄ and the solvent removed in vacuo leaving behind a clear, yellowish oil which solidified on standing. The crude products were purified by either recrystallization or column chromatography.
- 3-Benzyloxy-17β-methoxyestra-1, 3,5(10)-triene (3a). Recrystallization from methanol, 63% yield. Yellowish solid, m.p. 92-94°C; TLC R_f 0.83; H-NMR (CDCl₃) δ: 7.32-7.48 (m, 5H), 7.22 (dd, J=8.7 and J=2.10 Hz, 1H), 6.80 (d, J=2.4, 1H), 5.05 (s, 2H), 3.39 (s, 3H), 3.33 (t; 1H, J=8.7), 2.83 (m, 2H), 1.22-2.34 (m, 13H), 0.80 (s, 3H). MS: m/z 377 [M+H]⁺.
- 3-Benzyloxy-17β-ethoxyyestra-1, 3,5(10)-triene (3b). Column chromatography, 49 % yield. TLC R_f 0.71; ¹H-NMR (CDC1₃) δ: 7.45-7.30 (m, 5H), 6.79 (dd, J=8.7 and J=2.10 Hz, 1H), 6.71 (d, J=2.5, 1H), 5.02 (s, 2H), 3.55 (dq, J= 6.9 Hz and 2.1 Hz, 1H), 3.48 (dq, J= 7.0 Hz and 2.1 Hz, 1H), 3.39 (t, J=8.1 Hz, 1H), 2.84-2.81 (m, 2H), 2.31-1.37 (m, 13H), 1.18 (t, J=6.9 Hz, 3H), 0.79 (s, 3H). MS: *m/z* 391 [M+H]⁺.
- 3-Benzy [oxy-175-propoxyestra-1, 3,5(10)-triene (3c). Column chromatography. Yield.
 54%. White solid. TLC R_f 0.68, ¹H-NMR (CDC1₃) δ: 7.44-7.37 (m, 5H), 6.75 (dd, J=8.6 and J=2.1 Hz, 1H), 6.70 (d, J=2.7, 1H), 5.02 (s, 2H), 3.41 (dt, J= 6.9 Hz and 2.4 Hz, 2H), 3.37 (t, J=8.4 Hz, 1H), 2.84-2.81 (m, 2H), 2.31-1.37 (m, 15), 0.92 (t, J=6.6 Hz, 3H), 0.79 (s, 3H).
 MS: *m/z* 405 [M+H]⁺.
 - **3-Benzyloxy-170-butoxyestra-1, 3,5(10)-triene (3d).** Column chromatography, yield 52%.
- White solid. TLC R_f 0.65, 1 H-NMR (CDC1₃) δ: 7.45-7.30 (m, 5H), 6.79 (dd, J=8.7 and J=2.10 Hz, 1H), 6.71 (d, J=2.5, 1H), 5.02 (s, 2H), 3.55 (dq, J= 6.9 Hz and 2.1 Hz, 1H), 3.48 (dq, J= 7.0 Hz and 2.1 Hz, 1H), 3.39 (t, J=8.1 Hz, 1H), 2.84-2.81 (m, 2H), 2.31-1.37 (m, 13H), 1.18 (t, J=6.9 Hz, 3H), 0.79 (s, 3H). MS: m/z 419 [M+H]⁺.
- 3-Benzyloxy-17β-hexyloxyestra-1, 3,5(10)-triene (3e). Column chromatography, yield 63%. White solid. TLC R_f 0.75, ¹H-NMR (CDC1₃) δ: 7.49-7.34 (m, 5H), 6.74 (dd, J=8.7 and J=2.7 Hz, 1H), 6.71 (d, J=2.7,1H), 4.98 (s, 2H), 3.44 (dt, J=7.6Hz and 2.7 Hz, 2H) 3.36 (t, J=8.1 Hz, 1H), 3.55 (dq, J= 6.9 Hz and 2.1 Hz, 1H), 3.48 (dq, J= 7.0 Hz and 2.1 Hz, 1H), 3.39 (t, J=8.1 Hz, 1H), 2.84-2.81 (m, 2H), 2.31-1.37 (m, 13H), 1.18 (t, J=6.9 Hz, 3H), 0.79 (s, 3H). MS: *m/z* 447 [M+H]⁺.
- 3-Benzyloxy-17β-actyloxyestra-1, 3,5(10)-triene (3f). Column chromatography, 55% yield, yellow oil. TLC R_f 0.85, 1 H-NUR (CDC1₃) δ: 7.45-7.30 (m, 5H), 6.79 (dd, J=8.7 and J=2.10 Hz, 1H), 6.71 (1 H, J=7.7), 5.02 (s, 2H), 3.55 (dq, J= 6.9 Hz and 2.1 Hz, 1H), 3.48 (dq, J= 7.0 Hz and 2.1 Hz, 1H), 3.39 (t, J=8.1 Hz, 1H), 2.84-2.81 (m, 2H), 2.31-1.37 (m, 13H), 1.18 (t, J=6.9 Hz, 3H), 0.79 (s, 3H). MS: m/z 475 [M+H]⁺.

General Procedure for the Preparation of 17β-alkoxyestra-1, 3,5(10)-triene (4a-f). To a solution of 2.0 mmol 3a-f in 10mL of methanol was added 0.2 g of Pd/C (10%) and ammonium formate (1.00 g, 16mmol). The reaction mixture was stirred at room temperature for 1 hr. Then the Pd/C was then removed by filtration and solvent was 5 removed in vacuo. To the oily residue water was added and the resulting solid was collected by filtration. Either recrystallization or column chromatography was used for purification. 17β-Methoxyestra-1, 3,5(10)-trien-3-ol (4a). Recrystallization from methanol, 50% yield. White solid, m.p. 242-244°C; TLC: $R_f 0.48$; ¹H-NMR (DMSO) δ : 7.05 (d, J=8.40 Hz, 1H), 6.51(dd, J=8.40 Hz and 2.10 Hz, 1H), 6.45 (d, J= 2.40 Hz, 1H), 3.30 (s, 3H), 3.28 (t, j=8.25 Hz, 1H); 2.73-2.72 (m, 3H); 2.56-2.50 (m, 1H); 2.30-1.22 (m, 13H); 0.74 (s, 3H). ¹³C-NMR 10 (DMSO) δ: 156.7, 139.3, 132.7, 128.0,116.8,114.5, 92.2, 58.7, 51.7, 45.6, 44.6, 40.2, 39.8, 31.1, 29.2, 28.8, 28.1, 24.4, 13.6; MS: m/z 287 [M+H]⁺, 255 [M-OCH₃]⁺. Anal. C, H. 17β-Ethoxyestra-1, 3,5(10)-trien-3-ol (4b). Recrystallization from methanol, 50% yield, white solid; TLC: $R_f 0.57$; ¹H-NMR (CDC1₃) δ : 7.08 (d, J=8.7 Hz, 1H), 6.55 (dd, J=8.4 Hz, 2.1 Hz, 1H), 6.48 (d, J=2.4 Hz, 1H), 3.65 (qd, J=7.02 Hz and 2.48 Hz, 1H), 3.56 (qd, J= 15 7.05 Hz and 2.48 Hz, 1H), 3.44 (t, J=8.4 Hz, 1H), 2.76-2.72 (m, 2H), 2.20-1.10 (m, 13H), 1.20 (t, J=7.2 Hz, 3H), 0.80 (s, 3H); ¹³C-NMR (CDC1₃) δ: 155.72,138.83, 132.64, 127.19, 116.1, 113.7, 89.8, 66.1, 50.8, 44.5, 43.8, 39.3, 38.6, 30.1, 28.6, 27.8, 27.01, 23.5, 15.8 11.9; MS: m/z 301 [M+H]⁺, 255 [M-OC₂H₅]⁺. Anal. C, H. 17β-Propoxyestra-1, 3,5(10)-trien-3-ol (4c). Recrystallization from methanol, 50% yield, 20 white solid; TLC: $R_f 0.54$; ¹H-NMR (CDC1₃) δ : 7.08 (d, J=8.7 Hz, 1H), 6.55 (dd, J=8.4 Hz, 2.1 Hz, 1H), 6.48 (d, J=2.4 Hz, 1H), 3.45 (dt, J=6.77 Hz and 1.67 Hz, 2H), 3.31 (m, 3H), 2.76-2.72 (m, 2H), 2.20-1.10 (m, 13H), 0.94 (td, J=7.2 Hz and 1.92 Hz, 3H), 0.72 (s, 3H); ¹³C-NMR (CHC₁₃) δ: 154.0, 137.9, 131.7, 126.2, 115.0, 112.5, 89.0, 71.9, 50.1, 43.8, 43.2, $38.5, 38.0, 29.5, 27.9, 27.1, 26.3, 23.1, 22.9, 11.4, 10.4; MS: m/z 315 [M+H]^+, 255$ 25 $[M-OC_3H_7]^+$. Anal. C, H. 17β-Butoxyestra-1, 3,5(10)-trien-3-ol (4d). Recrystallization from methanol, 50% yield, white solid, m.p. 77-81°C; TLC: $R_f 0.47$; ¹H-NMR (CDCl₃) δ : 7.08 (d, J=8.7 Hz, 1H), 6.55 (dd, J=8.4 Hz, 2.1 Hz, 1H), 6.48 (d, J=2.4 Hz, 1H), 3.50 (dqn, J=7.00 Hz and 2.01 Hz, 1H), 3.45 (dqn, J=7.11 Hz and 1.85 Hz, IH), 3.31 (t, J=8.4 Hz, 1H), 2.76-2.72 (m, 2H), 2.20-1.10 30 (rn, 17H), 0.85 (t, J=7.2 Hz, 3H), 0.72 (s, 3H); 13 C-NMR (CHCl₃) δ : 153.3, 138.3, 132.7, 126.5, 115.2, 112.5, 89.1, 70.0, 50.3, 43.9, 43.3, 38.6, 38.1, 32.3, 29.6, 28.2, 27.1, 26.5, 23.0, 19.4, 14.0, 11.6; MS: m/z 329 [M+H]⁺, 255 [M-OC₄H₉]⁺.

17β-Hexyloxyestra-1, 3,5(10)-trien-3-ol (4e). Column chromatography, 70% yield, white semisolid. TLC: R_f 0.47; ¹H-NMR (CDCl₃) δ: 7.12 (d, J=8.4 Hz, 1H), 6.62 (dd, J=8.3 Hz, 2.7 Hz, 1H), 6.54 (d, J=2.5 Hz, 1H), 3.43 (dt, J=7.6Hz and 2.7 Hz, 2H) 3.36 (t, J=8.1 Hz, 1H), 2.80-2.77 (m, 2H), 2.25-1.25 (m, 18H), 0.89-0.85 (m, 6H), 0.78 (s, 3H); ¹³C-NMR (CHCl₃) δ: 153.2, 138.2, 132.6, 126.4, 115.1, 112.5, 89.0, 70.3, 50.2, 43.8, 43.3, 38.5, 38.0, 31.6, 30.1, 29.5, 28.1, 26.5, 25.8, 23.0, 22.6, 14.0, 11.6; MS: m/z 357 [M+H]⁺, 255 [M-OC₆H₁₃]⁺. Anal. C, H.

17β-Octyloxyestra-1, 3,5(10)-trien-3-ol (4f). Column chromatography, 75% yield, pale

yellow semi-solid. TLC: R_f 0.50; 1 H-NMR (CDC1₃) δ : 7.12 (d, J=8.7 Hz, 1H), 6.62 (dd, J=8.4 Hz, 2.2 Hz, 1H), 6.53 (d, J=2.3 Hz, 1H), 3.49 (qd, J=6.79 Hz and 2.52 Hz, 1H), 4.31 (qd, J=6.72 Hz and 2.55 Hz, 1H), 3.37 (t, J=8.5 Hz, 1H), 2.81-2.76 (m, 2H), 2.22-1.18 (m, 22H), 0.87-0.83 (m, 6H), 0.79 (s, 3H); 13 C-NMR (CHCl₃) δ : 153.3, 138.2, 132.6, 126.5, 115.2, 112.6, 89.1, 70.3, 50.2, 43.9, 43.3, 38.6, 38.0, 31.8, 30.1, 29.7, 29.4, 29.3, 28.1, 27.1,

26.4, 26.2, 23.0, 22.6, 14.0, 11.6; MS: m/z 385 [M+H]⁺, 255 [M-OC₈H₁₇]⁺. Anal. C, H.

- General Procedure for the Preparation of 3-Alkoxyestra-1, 3,5(10)-triene (5b,c). To compound 1 (0.5g, 1.8 mmol) and potassium carbonate (1.00g, 7.2.mmol) in 5 ml of acetone 10 mmol of 1-bromobutane or 1-bromooctane was added. The mixture was refluxed overnight then allowed to cool down and was filtered. The acetone was removed and the oily residue was purified.
- 3-Butoxyestra-1, 3,5(10)-trien-17β-ol (5b). Recrystallization from methanol: water 1:1 (v/v), 68% yield. White solid; m.p. 86-88°C; TLC R_f 0.62; ¹H-NMR (CDC1₃) δ: 7.17 (d, J=8.7 Hz, 1H), 6.70 (dd, J=8.4 Hz and 2.40 HZ, 1H), 6.62 (d, J=2.4 Hz, 1H), 3.93 (t, J=6.30 Hz, 2H), 3.71 (t, J=8.1 Hz, 1H), 2.86-2.80 (m, 2H), 2.20-1.10 (m, 17H), 0.96 (t, J=7.2 Hz, 3H), 0.77 (s, 3H); ¹³CNMR (CHCl₃) δ: 156.9, 137.7, 132.3, 126.1, 114.4, 111.9, 81.7, 67.5, 49.9, 43.8, 43.1, 38.7, 36.6, 31.3, 30.4, 29.7, 27.2, 26.3, 23.0, 19.2, 13.7, 10.9. MS: m/z 311 [M-OH]⁺.
- **3-Octyloxyestra-1, 3,5(10)-trien-17β-ol (5c).** Column chromatography, 72 % yield. White solid, m.p. 64-66°C; TLC R_f 0.70; H-NMR (CDC1₃) δ: 7.18 (d, J=8.7 Hz, 1H), 6.71(dd, J=8.7 Hz and 2.7 Hz, 1H), 6.62 (d, J=2.8 Hz, 114), 3.91 (t, J=6.6 Hz, 2H), 3.73 (t, J=8.4 Hz, 1H), 2.85-2.82 (m, 2H), 2.20-1.10 (m, 25 H), 0.88 (t, J=6.6 Hz, 3H), 0.77 (s, 3H); C-NMR (CHCI₃) δ: 156.9, 137.8, 132.4, 126.2, 114.5, 112.0, 81.9, 70.3, 67.9, 50.0, 43.9, 43.2, 38.8, 38.1, 36.6, 30.1, 29.7,29.4, 29.2, 27.2, 26.4, 26.2, 23.1, 22.6, 14.0, 11.0. MS: *m/z* 368 [M-OH]⁺. Anal. C, H.

Table 1. Crystal data and structure refinement for 4d.

Identification code 4d C23 H36 O3 Empirical formula Formula weight 360.52 Temperature 173(2) K Wavelength 0.71073 Å Crystal system Monoclinic Space group P2(1) Unit cell dimensions a = 8.6418(4) Å $\alpha = 90^{\circ}$. b = 9.5698(5) Å $\beta = 102.021(1)^{\circ}$. c = 12.8534(7) Å $\gamma = 90^{\circ}$. $1039.67(9) \text{ Å}^3$ Volume 1.152 Mg/m^3 Density (calculated) 0.074 mm⁻¹ Absorption coefficient F (000) 396 Crystal size $0.21 \times 0.21 \times .13 \text{ mm}^3$ Theta range for data collection 1.62 to 27.50°. Index ranges -11• h• 11, -12• k• 8, -16• 1• 16 Reflections collected 7032 Independent reflections 3784 [R (int) = 0.0233]Completeness to theta = 27.49° 99.8% Absorption correction Integration Max. and min. transmission 0.996 and 0.987 Full-matrix least-squares on F² Refinement method Data / restraints / parameters 3784 / 1 / 247 Goodness-of-fit on F² 0.976 Final R indices [I>2sigma(I)] R1 = 0.0371, wR2 = 0.0890 [3294] R indices (all data) R1 = 0.0434, wR2 = 0.0917Absolute structure parameter -0.6(10)Extinction coefficient 0.007(2)0.205 and -0.172 e. Å⁻³ Largest diff. peak and hole $RI = \Sigma(IIF_oI - IF_cII) / \Sigma IF_oI$ $wR2 = [\sum [w(F_o^2 - F_c^2)^2] / \sum [w(F_o^2)^2]]^{1/2}$ $S = \left[\sum \left[w(F_0^2 - F_c^2)^2\right] / (n-P)\right]^{1/2}w - \frac{1}{[o^2(F_0^2) + (0.0370*p)^2 + 0.31*p]}, p = \left[\max(F_0^2, 0) + 2*; F_c^2\right]/3$

Example 2: Biological activity of compounds

Cytotoxicity Studies. Mouse clonal hippocampal HT-22 cells were cultured in DMEM media supplemented with 10% fetal bovine serum under standard cell culture conditions. All wells in the 96 well culture plate contained approximately 5,000 HT-22 cells as determined by a Neubauer hemacytometer and the cells were incubated for 24 hrs before the compounds were added. The estradiol derivatives were purified recrystallization or column chromatography and were free from (1) as determined by HPLC. All agents were dissolved in absolute ethanol and diluted, with the culture media, to a final concentration of 0.01 μM; 0.1 μM; 1.0 μM; and 10 μM in their respective wells. The cells were further incubated for 24 hrs before sodium glutamate in a solution of phosphate buffer was added. Cell viability was quantified 2 hrs later by the calcein AM assay (Green, P.S., E.J.Perez, T. Calloway and J.W. Simpkins: (2000), Journal of Neurocytology, Vol. 29, pp. 419-423) in a phosphate buffer solution.

Statistical Analysis. ANOVA was used to determine the significance of differences among groups. Comparison between groups were was done using the Tukey test. Ap < 0.05 was considered significant. The results are shown in Figure 4.

Compared to (1), 4c-f of the six 17β -O-alkylestradiols showed improved neuroprotection in a dose-dependent manner against the glutamate- induced oxidative damage in murine HT-22 cells at concentrations of 0.1 μ M and higher (Fig. 4). These compounds were essentially equipotent at 1 μ M (approximately twice as many cells were viable compared to the control), and showed no apparent relationship with a single molecular property such as lipophilicity (based on the calculated log P). The logarithm of the 1-octanol/water partition coefficient (log P) was calculated by an atom fragment method implemented in the molecular modeling package HyperChem version 6.0 (Hypercube, Gainesville, FL): Ghose, et al., (1988) J. Comput Chem, Vol. 9, pp. 80-90. The obtained log P values were as follows: 4.01 (1), 4.29 (4a), 4.63 (4b), 5.10 (4c), 5.49 (4d), 6.29 (4e), and 7.08 (4f). The calculated log P for the 3-alkylestradiols were 4.09 (5a), 5.25 (5b), and 6.83 (5c).

The butyl (4e) and octyl ether (4f) were neuroprotective to a similar extent at a concentration of 10 μ M and 1 μ M. In contrast, the parent compound (1) and 17 β -methylestradiol were effective only at 10 μ M, and were less active then 4c and 4e at this

concentration. 17β -ethylestradiol (4a) was ineffective even at $10 \,\mu\text{M}$. The 5(b) and 5c ethers in which the phenolic hydroxyl in the A-ring were blocked were ineffective with respect to cytoprotection.

The complex relationship of cytoprotection and 17-alkoxy chain length was surprising. A comparison of the solid-state conformation of 4a and 4d revealed no apparent differences in the preferred geometry of the steroid backbone between a representative "active" (4e) and an "inactive" (4a) ether derivative of (1). Without wishing to be limited by theories, we propose that a possible explanation for the above described behavior is that the interaction of the alkyl chain of the $17(\beta)$ -substituent with the target site or the lipoidal cell membrane plays an important role in the efficacy of the derivative as a cytoprotectant. Thus, 4a and 4b having a compact alkyl group may not have the flexibility (i.e., sufficient degrees of freedom for bond rotation) to embed into a cell membrane effectively; however, a longer alkyl chain (C \geq 3) may provide this property.

In summary, 17β and 17α -alkyl ethers of estradiol have dose-dependent cytoprotective effects *in vitro*. Moreover, this effect is manifested at lower concentration ($<1\mu M$) than that of the parent compound.

Example 3: Cytoprotection (neuroprotection) is unrelated to binding to estrogen receptor

Human cloned estrogen receptors (ER) for both ER α and ER β areas were mixed with radiolabeled 17 β -estradiol and with no other compound (total binding), with excessive amount of diethylstilbesterol (non-specific binding), or with cold (unlabeled) estradiol, or the test compound. All groups were determined in duplicate or triplicate. 17 β -estradiol was tested at concentrations of 0.1, 1 and 10 mM, while all other test compounds were assayed at 10 mM.

 17β -estradiol produced a dose-dependent inhibition of binding of the labeled estradiol to both receptors with approximately equal affinity. The activity of 17β estradiol was assigned a value of 1. Test compounds were compared to the binding inhibition produced by 17β -estradiol.

Values of < 0.01 indicate no evidence of binding of the test compound to the receptor. Values of < 0.1 indicate weak binding (less than 10% of the activity of 17 β -estradiol.

ND indicates that the compound has not been tested at this time

Table 2: Comparison of compounds based on neuroprotective properties and estrogen receptor binding.

COMPOSITE	NEUROPROTECTION (Effectiveness relative to E2)	ERα BINDING (Relative to E2)	ERβ BINDING (Relative to E2)
17beta E2	1	1	1
Ent-E2	1.14117	<0.028	<0.028
17alpha E2	1.35856	ND	ND
17-ethyl ether		<0.01	ND
17-octyl ether		<0.01	<0.01
17-propyl ether		<0.01	ND

Although certain preferred embodiments of the present invention have been described, the spirit and scope of the invention is by no means restricted to what is described above. In addition to the above references incorporated by reference, Prokai et al. (2001) J. Med. Chem. 2001, Vol 44, 110-114 is also incorporated by reference.